TWO LABDANE DITERPENOIDS FROM NICOTIANA RAIMONDII

MASANA NOMA, FUMIYO SUZUKI, KENJI GAMOU and NOBUMARO KAWASHIMA

Central Research Institute, The Japan Tobacco and Salt Public Corporation, 6-2 Umegaoka, Midoriku, Yokohama, Kanagawa 227, Japan

(Revised received 5 June 1981)

Key Word Index-Nicotiana raimondii; Solanaceae; new diterpene; labdanoids.

Abstract—Two new labdanoids from *Nicotiana raimondii* have been identified as (12E) - 7β - hydroxylabda - 8(20), 12, 14 - triene and 16 - hydroxy - labda - 8(20), 13(16), 14 - triene.

INTRODUCTION

The diterpenes in exudates from trichomes of leaves of all species of *Nicotiana* examined so far are species specific and can be used as chemotaxonomic markers [1,2]. This finding has led us to investigate the diterpenes in exudates among various species of *Nicotiana*. *Nicotiana* raimondii (Paniculatae) exudes especially large amounts of gummy materials which sometime reach up to 0.3% of the dry wt of green leaves. We now report on the main components of this exudate.

RESULTS AND DISCUSSION

The 13 C NMR spectra of compounds 1 and 2 (named raimonol and isoraimonol, respectively) and related compounds are shown in Table 1. The presence in 1 and 2 of the A ring moiety including three Me groups found in 3, obtained as a fungal metabolite of *cis*-abienol [3], is indicated by the fact that their spectra contain the same signals as those assigned to C-1, C-2, C-3, C-4, C-17, C-18 and C-19 of 3. In 1 three of the signals (δ 11.9, 110.0 and 141.6) are as assigned to C-14, C-15 and C-16 of *trans*-

Table 1. ¹³C NMR spectra of compounds 1-5 (25.0 MHz, CDCl₃, TMS as internal standard)

	Raimonol	Isoraimonol (2)	(2)	(4)	t-Abienol (5)
	(1)		(3)		
C-1	39.1 t	39.2 t	39.3 t	39.1 t	40.1 t
C-2	19.4 t	19.3 t	19.5 t	19.3 t	18.6 t
C-3	42.0 t	42.0 t	42.2 t	42.1 t	41.9 t
C-4	33.6 s	33.9 s	33.5 s	33.5 s	33.2 s
C-5	53.1 d	53.2 d	55.5 d	55.3 d	56.1 d
C-6	33.4 t	33.4 t	24.3 t	24.1 t	20.4 t
C-7	73.8 d	73.8 d	38.2 t	38.0 t	44.1 t
C-8	150.5 s	150.4 s	148.2 s	148.5 s	73.7 s
C-9	55.3 d	54.8 d	57.4 d	57.0 d	62.2 d
C-10	39.3 s	38.8 s	39.6 s	39.4 s	38.9 s
C-11	22.9 t	21.1 t	22.1 t	23.0 t	24.0 t
C-12	133.3 d	30.0 t	133.9 d	134.0 d	136.4 d
C-13	133.7 s	146.8 s	131.4 s	133.2 s	132.1 s
C-14	141.6 d	139.0 d	131.6 d	141.6 d	141.7 d
C-15	110.0 t	113.1 t	113.0 t	109.6 t	110.0 t
C-16	11.9 q	115.5 t	19.7 q	11.7 q	11.8 <i>q</i>
C-17	14.5 q	14.5 q	14.5 q	$14.3 \; q$	15.4 q
C-18	33.6 q	33.5 q	33.6 q	33.5 q	33.5 q
C-19	21.7 q	21.6 q	21.8 q	21.6 q	21.6 q
C-20	104.2 t	$103.1 \ t$	107.7 t	107.3 t	24.1 q

396 M. Noma et al.

abienol (5), and this together with its UV absorption (230 nm, log 4.36) shows the presence of a trans diene moiety. Changing the C-16 Me group (δ 11.9, q) of raimonol to a terminal methylene (δ 115.5, t) affords isoraimonol. The ¹H NMR data of 1 and 2 are shown in Table 2. In double resonance experiments, irradiation at δ 3.99 (1H, q, J = 5.5, 11 Hz) changed an octet (1H, J = 2.5, 5.5 and 12 Hz) at δ 2.09 and a sextet (1H, J = 12, 12 and 11 Hz) at δ 1.28 to a broad doublet and a triplet respectively and resulted in the elongation of singlets at δ 4.67 and 5.19. These changes, together with the presence of a quartet (1H, J = 2.5, 12 Hz) at δ 1.16, point to the presence of the partial structure A in 1 and 2 (Fig. 1). The configurations of the protons Ha, Hb, Hc and Hd which satisfy the Karplus rule are equatorial, axial, equatorial and axial, respectively. Thus the hydroxyl group is in an equatorial position. In a triple resonance experiment with 1, irradiation at a triplet (1H, J = 6.5 Hz) at δ 5.42 and a broad doublet at δ 1.70 changed a septet (1H, J = 6.5, 11 and 16 Hz) at $\delta 2.24$ and an octet (1H, 1)J = 2.5, 6.5 and 16 Hz) at δ 2.42 to AB doublets (J = 16 Hz) affording the partial structure **B**. Tosylation of 1 followed by reduction with LiAlH₄ gave 4 which was identical (IR and MS) with the compound obtained by dehydration of 5. This confirmatory evidence and the positive specific rotation of these hydrocarbons established the structures of raimonol (1) and isoraimonol (2) as $(12E) - 7\beta$ - hydroxylabda -8(20),12,14 - triene and 7β - hydroxylabda -8(20),13(16),14 - triene.

Table 2. ¹H NMR spectra of raimonol (1) and isoraimonol (2) (400 MHz, CDCl₃, TMS is internal standard)

	Raimonol	Isoraimono
H-5	1.16 (1H, q)	1.16 (1H, q)
H-6 ax	1.28 (1H, sextet)	1.28 (1H, sextet)
H-6 eq	2.09 (1H, octet)	2.12 (1H, octet)
H-7	3.99(1H, q)	$4.00(1\mathrm{H},q)$
H-9	1.70(1H,br,d)	1.43(1H,br,d)
H-11	2.24 (1H, septet)	*
	2.42 (1H, octet)	*
H-12	5.42(1H, t)	*
H-14	6.35(1H,q)	6.39(1H,q)
H-15	4.91(1H,d)	5.24(1H,d)
	5.07(1H, d)	5.08(1H, d)
H- 16	1.76(3H, s)	5.00(1H, s)
		5.03(1H, s)
H-17	0.92 (3H, s)	0.91(3H, s)
H-18	* 0.84 (3H, s)	*0.81(3H, s)
H-19	0.72(3H, s)	0.68(3H, s)
H-20	4.67 (1H, s)	4.76(1H, s)
	5.19(1H, s)	5.23(1H, s)

^{*}Unassigned.

isothermal). R_t s of 1 and 2 were 5.0 and 4.0 min, respectively. 180 mg 1 and 60 mg 2 could have been obtained from the entire fraction on the basis of GLC analysis, but these compounds were purified by prep. GLC as required, because

EXPERIMENTAL

Isolation of raimonol (1) and isoraimonol (2). Exudate of field-grown N. raimondii was washed out from green leaves (1 kg) by soaking in CHCl₃ for 30 sec (\times 2), and purified by SiO₂ column. The Et₂O-hexane (1:4) fraction (400 mg), after washing with hexane, was subjected to prep. GLC (1 m \times 3 mm, packed with 5% 0V-101, He 50 ml/min, 240°

they are not very stable when purified. Reinjection of pure 1 or 2 gave a single peak at the expected R_i . Therefore, these isomers are not products of thermal isomerization.

Raimonol. Mp 101–103° (fine needles); $[\alpha]_D^{25} + 24.8$ ° (c = 0.55, EtOH); UV $\lambda_{\text{max}}^{\text{EtOH}}$ 230 nm (log ϵ 4.36); IR ν_{max} cm⁻¹: 3250, 3040, 1640, 1604, 1100, 1062, 1038, 990; MS (low resolution) m/z: 288 [M]⁺, 231, 206, 185, 151, 123.

Fig. 1. Partial structures A and B. Numbers above or below H are chemical shifts of the proton in ¹H NMR. Numbers between arrows are coupling constants between the protons indicated by the arrows.

В

Isoraimonol. Mp 70–72° (needles); $[\alpha]_{25}^{25} + 18.1°$ (c = 0.24, EtOH); UV $\lambda_{\text{max}}^{\text{EtOH}}$ 223 nm (shoulder) (log ϵ 3.52); IR ν_{max} cm⁻¹: 3400, 3100, 1650, 1208, 1100, 1038, 900; MS (low resolution) m/z: 288 (M⁺), 273, 255, 206, 135, 121.

Hydrocarbon (4) from cis-abienol. To a soln of transabienol (1.3 g) [obtained from cis-abienol [4] isolated from Canadian fir balsam (Abies balsameae)] in pyridine (26 ml), POCl₃ (910 μ l) was added dropwise at 4°. After 48 hr at 4°, the reaction mixture was poured into ice H₂O and extracted with CH₂Cl₂. Purification on SiO₂ containing AgNO₃ (25%, w/w) (hexane as an eluant) gave 4 (436 mg), $[\alpha]_{365}^{125} + 77.0^{\circ}$

 $(c = 0.693, \text{ CCl}_4)$ $[\alpha]_{546}^{256} + 31.8^{\circ}; \text{ IR } \nu_{\text{max}} \text{ cm}^{-1}; 3090, 1640, 1602, 1200, 1085, 985, 890; MS (low resolution) } m/z: 272 [M]^+, 257, 216, 201, 191, 135; <math>^{1}\text{H NMR}$ (100MHz, CCl₄, TMS as int. standard): δ 6.20 (1H, q), 5.30 (1H, t), 4.94 (1H, d), 4.84 (1H, d), 4.76 (1H, s), 4.43 (1H, s), 1.65 (3H, s), 0.87 (3H, s), 0.82 (3H, s), 0.72 (3H, s).

Hydrocarbon 4 from 1. To a soln of 1 (10 mg) in pyridine (0.2 ml), TsCl (67 mg) in pyridine (0.3 ml) was added dropwise with cooling by ice. After 16 hr at room temp., the tosylate (6 mg) was obtained by usual work-up. ¹H NMR (100 MHz, CCl₄, TMS as int. standard): δ 7.77 (2H, d), 7.28 (2H, d), 6.20 (1H, q), 5.08 (1H, d), 5.04 (s), 4.83 (1H, d), 4.52 (s), 2.44 (3H, s), 1.68 (3H, s), 0.86 (3H, s) 0.80 (3H, s), 0.66 (3H, s). Without further purification, the tosylate was reduced with LiAlH₄ (2.6 mg) in boiling Et₂O (2 ml). After 2 hr, the reaction mixture was passed through a small SiO₂ column (200 mg), and the eluate (2 mg) purified by prep. GLC (1 m×3 mm, packed with 5% OV-101, temp. programmed 160-250° at 2°/min, He 50 ml/min) to give the hydrocarbon (400 µg): IR and MS spectra identical with those from cis-abienol; $[\alpha]_{365}^{25}$ ca + 49°, $[\alpha]_{365}^{25}$ ca + 12° (c = ca 0.016, CCl₄).

Acknowledgements—We thank Mr. T. Hieda, this institute, for his kind gift of cis-abienol, and Dr. T. Yokota and Mr. K. Furihata, University of Tokyo, for the measurement of ¹H NMR spectra.

REFERENCES

- Reid, W. W. (1979) in The Biology and Taxonomy of the Solanaceae (Hawkers, J. G., Lester, R. N. and Skelding, A. D., eds), Linnean Society Symposium Series. No. 7, pp. 273-278.
- Tomita, H., Sato, M. and Kawashima, N. (1980) Agric. Biol. Chem. 44, 2517.
- 3. Hieda, T., Mikami, Y., Obi, Y. and Kisaki, T. (1980) Proc. Annu. Meeting Soc. Ferm. Tech. (Japan), p. 57.
- 4. Mills, J. S. (1967) J. Chem. Soc. C 2514.